

Teaching cardiac electrophysiology modeling to undergraduate students: laboratory exercises and GPU programming for the study of arrhythmias and spiral wave dynamics

Ezio Bartocci, Rupinder Singh, Frederick B. von Stein, Avestia Amedome, Alan Joseph J. Caceres, Juan Castillo, Evan Closser, Gabriel Deards, Andriy Goltsev, Roumwelle Sta. Ines, Cem Isbilir, Joan K. Marc, Diqun Moore, Dana Pardi, Sandeep Sadhu, Samuel Sanchez, Pooja Sharma, Anoop Singh, Joshua Rogers, Aron Wolinetz, Terri Grosso-Applewhite, Kai Zhao, Andrew B. Filipinski, Robert F. Gilmour, Jr., Radu Grosu, James Glimm, Scott A. Smolka, Elizabeth M. Cherry, Edmund M. Clarke, Nancy Griffith and Flavio H. Fenton

Advan in Physiol Edu 35:427-437, 2011. ;
doi: 10.1152/advan.00034.2011

You might find this additional info useful...

This article cites 67 articles, 11 of which you can access for free at:
<http://advan.physiology.org/content/35/4/427.full#ref-list-1>

Updated information and services including high resolution figures, can be found at:
<http://advan.physiology.org/content/35/4/427.full>

Additional material and information about *Advances in Physiology Education* can be found at:
<http://www.the-aps.org/publications/ajpadvan>

This information is current as of January 17, 2013.

Teaching cardiac electrophysiology modeling to undergraduate students: laboratory exercises and GPU programming for the study of arrhythmias and spiral wave dynamics

Ezio Bartocci,¹ Rupinder Singh,² Frederick B. von Stein,³ Avesse Amedome,⁴ Alan Joseph J. Caceres,⁴ Juan Castillo,⁴ Evan Closser,⁴ Gabriel Deards,⁴ Andriy Goltsev,⁴ Roumwelle Sta. Ines,⁴ Cem Isbilir,⁴ Joan K. Marc,⁴ Diqun Moore,⁴ Dana Pardi,⁴ Sandeep Sadhu,⁴ Samuel Sanchez,⁴ Pooja Sharma,⁴ Anoop Singh,⁴ Joshua Rogers,⁴ Aron Wolinetz,⁴ Terri Grosso-Applewhite,⁴ Kai Zhao,⁴ Andrew B. Filipski,⁵ Robert F. Gilmour, Jr.,³ Radu Grosu,⁶ James Glimm,¹ Scott A. Smolka,⁶ Elizabeth M. Cherry,^{3,7} Edmund M. Clarke,⁸ Nancy Griffith,⁴ and Flavio H. Fenton³

¹Department of Applied Mathematics and Statistics, Stony Brook University, Stony Brook; Departments of ²Biomedical Engineering and ³Biomedical Sciences, Cornell University, Ithaca; ⁴The City University of New York, New York;

⁵Department of Software Engineering, Rochester Institute of Technology, Rochester; ⁶Department of Computer Science, Stony Brook University, Stony Brook; ⁷School of Mathematical Sciences, Rochester Institute of Technology, Rochester, New York;

and ⁸Computer Science Department, Carnegie Mellon University, Pittsburgh, Pennsylvania

Submitted 26 April 2011; accepted in final form 1 September 2011

Bartocci E, Singh R, von Stein FB, Amedome A, Caceres AJ, Castillo J, Closser E, Deards G, Goltsev A, Ines RS, Isbilir C, Marc JK, Moore D, Pardi D, Sadhu S, Sanchez S, Sharma P, Singh A, Rogers J, Wolinetz A, Grosso-Applewhite T, Zhao K, Filipski AB, Gilmour RF Jr, Grosu R, Glimm J, Smolka SA, Cherry EM, Clarke EM, Griffith N, Fenton FH. Teaching cardiac electrophysiology modeling to undergraduate students: laboratory exercises and GPU programming for the study of arrhythmias and spiral wave dynamics. *Adv Physiol Educ* 35: 427–437, 2011; doi:10.1152/advan.00034.2011.— As part of a 3-wk intersession workshop funded by a National Science Foundation Expeditions in Computing award, 15 undergraduate students from the City University of New York¹ collaborated on a study aimed at characterizing the voltage dynamics and arrhythmogenic behavior of cardiac cells for a broad range of physiologically relevant conditions using an *in silico* model. The primary goal of the workshop was to cultivate student interest in computational modeling and analysis of complex systems by introducing them through lectures and laboratory activities to current research in cardiac modeling and by engaging them in a hands-on research experience. The success of the workshop lay in the exposure of the students to active researchers and experts in their fields, the use of hands-on activities to communicate important concepts, active engagement of the students in research, and explanations of the significance of results as the students generated them. The workshop content addressed how spiral waves of electrical activity are initiated in the heart and how different parameter values affect the dynamics of these reentrant waves. Spiral waves are clinically associated with tachycardia, when the waves remain stable, and with fibrillation, when the waves exhibit breakup. All *in silico* experiments were conducted by simulating a mathematical model of cardiac cells on graphics processing units instead of the standard central processing units of desktop computers. This approach decreased the run time for each simulation to almost real time, thereby allowing the students to quickly analyze and characterize the simulated arrhythmias. Results from these simulations, as well as some of the background and methodology taught during the workshop, is presented in this article along with the programming code and the explanations of simulation results in an effort to allow other teachers and students to perform their own demonstrations, simulations, and studies.

ventricular fibrillation; atrial fibrillation; graphics processing unit simulations

THE 2011 ATRIAL FIBRILLATION WORKSHOP was one of a series of annual workshops affiliated with the Computational Modeling and Analysis of Complex Systems (CMACS), a multi-institutional (8) and multi-principal investigator (19) project led by Edmund Clarke. CMACS is funded by a National Science Foundation Expeditions in Computing award. The objective of the workshops is to develop the scientific interest and skills of students from urban minority-serving institutions and especially to motivate them to study the kinds of computational modeling techniques and applications used and developed in the project. Each year, the workshop is held in January at Lehman College in the Bronx, New York. Students from all parts of The City University of New York (CUNY), in which Lehman College is only one of many colleges, are invited to apply for the workshop; applicants are admitted based on grades and recommendations. The first workshop was held in January 2010 and addressed the role of cellular signaling pathways in the development of pancreatic cancer. Instruction at the 2010 workshop was provided by faculty members from Lehman College (N. Griffith), the University of Pittsburgh (James Faeder), Carnegie Mellon (Christopher Langmead), and New York University (Bud Mishra) together with several CUNY graduate students (Loes Olde Loohuis, Ziping Liu, and Fred Dieckamp). For the 2011 workshop, the instructors were from Cornell University (F. H. Fenton and R. F. Gilmour, Jr.), Stony Brook (E. Bartocci, S. A. Smolka, and J. Glimm), and Lehman College (N. Griffith) together with several Cornell and CUNY graduate students (R. Singh, F. B. von Stein, T. Grosso-Applewhite, K. Zhao, J. Rogers, and A. Wolinetz).

The Workshop

The Atrial Fibrillation workshop consisted of three related week-long units, each with a series of lectures and laboratory exercises, conducted 5 days/wk for 5 h/day. The teaching philosophy of the workshops is to engage the students in discussion, group work, and activities more than

¹ Brooklyn College, Hunter College, Lehman College, and Queens College.
Address for reprint requests and other correspondence: F. H. Fenton, Dept. of Biomedical Sciences, Cornell Univ., T7012C Veterinary Research Tower, Ithaca, NY 14850 (e-mail: fhf3@cornell.edu).

lectures. The workshop was organized around alternating lectures with laboratories and group activities to implement this philosophy.

In the first week, F. H. Fenton and R. Singh gave a series of lectures and laboratory exercises on excitable systems in biology (41, 42, 66, 72), physics (39, 45, 55), chemistry (6, 8, 78), and mathematics (26, 43, 48). This background provided students with a robust understanding of excitable systems and their relationship to complex systems and chaos (35, 71, 77). Other material in the first week emphasized the role of both mathematical and computational frameworks in the study of cardiac arrhythmias (16, 31). The motivation for computational and mathematical approaches in the study of cardiac arrhythmias was further conveyed to the students by explaining how the field of cybernetics, founded by Norbert Wiener, was a result of his studies not only on anti-aircraft defense but also of cardiac arrhythmias such as fibrillation (75) at the beginning of the 1940s. Emphasis was given to inquiry-based teaching. As a result, for the first part of the week, students experimented with density (saline) oscillators (51) and chemical oscillators (6, 8, 52, 78) to relate the nature of the oscillations in each system with those of cardiac action potentials (APs) and to quantify in a simple way the similarities between the complex dynamics in these systems and those of cardiac arrhythmias. The second part of the week concentrated more on how the membrane potential of cells such as neurons and cardiomyocytes can be represented mathematically. Various models (26) with different levels of biophysical detail were simulated and studied using interactive Java applets (24) (created by E. M. Cherry, A. B. Filipski, and F. H. Fenton). At the end of the first week, R. F. Gilmour, Jr., gave a lecture on experimental methods that illustrated how the electrophysiological properties F. H. Fenton had introduced in the context of computational models could be observed and measured experimentally.

The second week began with an exercise (designed by T. Grosso-Applewhite) with two goals in mind. The first, overt goal was to provoke the students to think critically about the value of models and what characteristics make them valuable. The second, unstated goal was motivated by observations from the first workshop that some of the student groups tended to degenerate into three people working side by side instead of collaboratively. This goal was to acquaint the students with each other and to push them out of their usual mode of individual student work, encouraging them instead to contribute actively to the group's progress. The exercise asked students to first list the characteristics of good models, individually, and then to prioritize them as a group. Subsequent group work validated the success of this exercise.

N. Griffith and K. Zhao lectured on various numerical methods used to analyze the mathematical models presented during the first week, and J. Rogers presented a tutorial on Compute Unified Device Architecture (CUDA) programming (62). CUDA provides a programming interface for the highly parallel graphics chips (graphics processing units) from NVIDIA, which are now standard in many laptops and desktops. Because CUDA programming uses the multiprocessing capabilities of the NVIDIA chips (3, 62), it provides much faster calculations than those obtained using the central processing units. For the workshop, J. Rogers and A. Wolinetz (both Master's degree students in the Lehman College Computer Science program) made available two computers that they had built around

these high-performance NVIDIA graphics chips and supported their use by the undergraduate students. At the end of the second week, E. Bartocci (Stony Brook University) gave an in-depth series of lectures on the implementation of an electrophysiological model of a cardiac cell (9) in a CUDA program.

During the final week, E. Bartocci and the students modified the E. Bartocci, E. M. Cherry, and F. H. Fenton CUDA program to simulate two-dimensional idealized sections of cardiac tissue and to examine a set of five different parameter spaces in detail to identify how certain changes in physiology affect the stability and dynamics of reentrant waves (spiral waves). The students subsequently prepared presentations with their results and interpretations, which were given on the final day of the workshop. All lectures and exercises can be found at <http://www.lehman.edu/academics/cmecs/outline.php>.

The Saline Oscillator

As a first approach to introducing students to the concepts of nonlinear dynamics and chaos necessary to understand cardiac dynamics (35, 71), in general, and arrhythmias, in particular, from both a physical and mathematical point of view, F. H. Fenton and R. Singh prepared a series of exercises using several setups of saline oscillators. A density (in this case, saline) oscillator is a very simple and inexpensive, yet extremely interesting, system discovered by accident in the late 1960s by Martin Seelye (51), where a container with a higher-density fluid is immersed into another fluid with a lower density with a narrow orifice connecting the two containers (see Fig. 1A). The difference in densities, combined with gravity, leads to a Rayleigh-Taylor instability (37, 64) that results in a rhythmic change in flow of fluid from one container to the other that can last for many hours (12, 39, 58). Although the phase transition dynamics are complicated (59), qualitatively the denser fluid has a tendency to fall and the less dense fluid has a tendency to rise. Separating the two solutions via hydrostatic pressure alone is a difficult balancing act. Typically, a small amount of solution will escape in one direction or the other, creating a jet. Over some period of time, this jet eventually will taper off as too much liquid accumulates on the receiving container; the imbalance in hydrostatic pressure then drives a jet in the other direction. The phenomenon continues as long as the density difference across the orifice is large enough to sustain it. This flow or leak of saline water (when going from the upper chamber to the lower chamber) and fresh water (when going from the lower chamber to the upper chamber) is clearly visible (see the movie at <http://TheVirtualHeart.org/CMACS/salineoscillator.mov>) as a laminar flow through the orifice and as a buoyant jet at the orifice exits. Furthermore, the flux across the orifice can be easily quantified using two electrodes and an oscilloscope. By placing one electrode in line with the orifice and the other far away from it, oscillatory changes in the boundary layer of the electrode in line with the orifice can be used to measure a change in voltage of ~ 100 mV that quantifies the presence and absence of a jet (73). This oscillating voltage signal is not only qualitatively similar to an AP from an autooscillatory sinoatrial (SA) node cell but, in some respects, is also qualitatively similar to cardiomyocyte APs in general (see Fig. 1B). In both cardiac cells and the oscillator, it is possible to provide an initiating stimulus [a

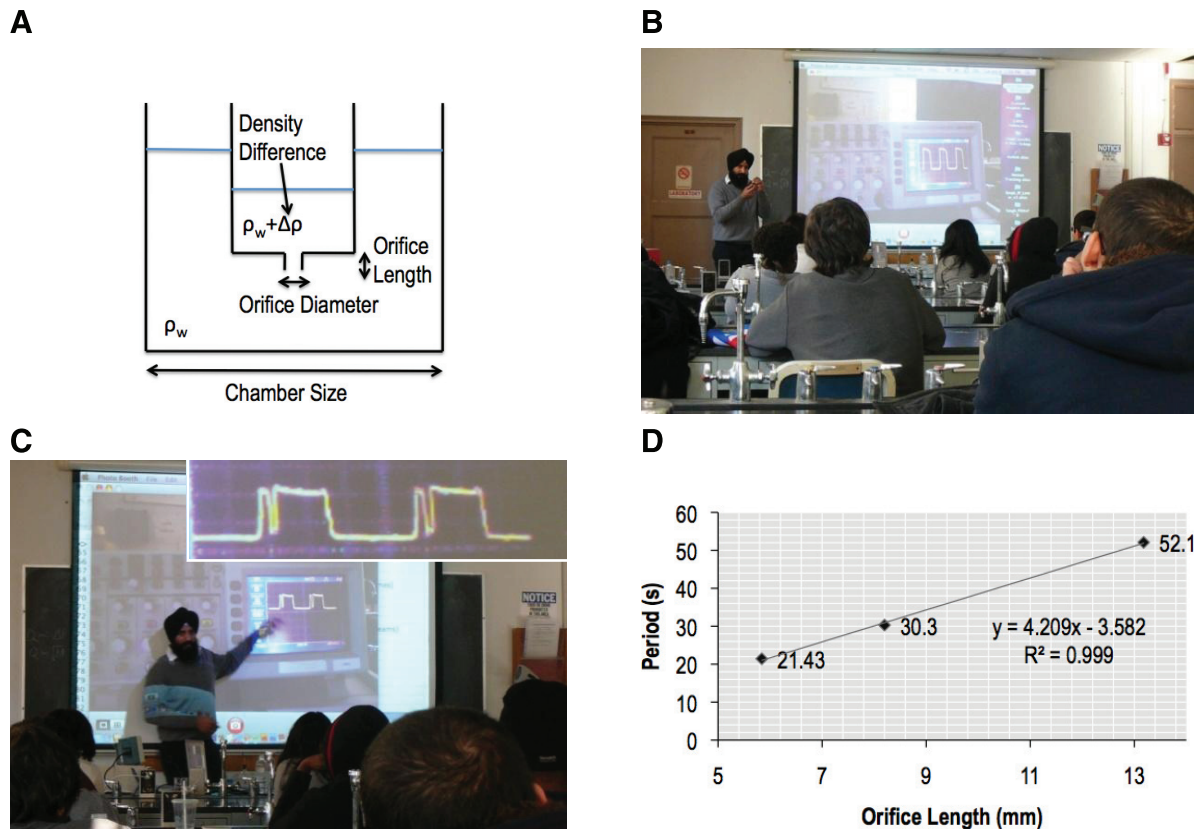


Fig. 1. Saline oscillator. *A*: schematic illustration for the setup. ρ_w , water density; $\Delta\rho$, density difference between saline and fresh water. *B*: demonstration of the voltage signal measured with an oscilloscope. *C*: demonstration of alternans during forced pacing. The *inset* shows an enlarged version of the image projected on the screen. *D*: example of the data obtained by students using the saline oscillator with different orifice lengths (see *A*). Note the linear trend in period of oscillation, consistent with the Hagen-Poiseuille law. (For a movie of the saline oscillator, see <http://thevirtualheart.org/CMACS/salineoscillator.mov>.)

small injection of fresh water in the outer container for the oscillator (39)], and, by constant periodic stimulation, it is possible to induce complex rhythms such as alternans (see Fig. 1C) (40, 60, 74) and Wenckebach rhythms (23, 34, 69). Therefore, the oscillator is an excellent system to provide students with a hands-on experience demonstrating how excitable systems behave and how the oscillator, in particular, shares many dynamic similarities with cardiac cells.

For the experiments, students were divided into four groups, each of which studied the effects of changing four physical parameters on the system's dynamics. In particular, each team tried several different orifice diameters, orifice lengths, density differences, and chamber sizes and recorded the changes in the oscillation period. After cycling through each of the four exercises, each team was asked to plot their results, compare their findings with those of the other groups, and supply an explanation for the observed trends based on their physical intuition. Afterward, a detailed explanation of the results was given in a lecture. Figure 1D shows one example in which the orifice length was changed. For the orifice length chosen in our setup, the time scales of the laminar flow were larger than the time scales corresponding to the transitions in the flow reversal; as a result, the flow rate through the orifice was assumed to be largely governed by the Hagen-Poiseuille law (44). An increase in orifice length led to a linear decrease in flow rate and, in turn, to an approximately linear increase in the period of oscillation.

Chemical Oscillators

Although the saline oscillator allowed the students to understand some of the concepts of complex systems that are also found in cardiac dynamics, such as oscillations between unexcited and excited states (polarization-depolarization) along with alternans and higher-order periodicities (Wenckebach rhythms), the dynamics were confined to a single cell or oscillator, so it was not possible to study spatial effects. Chemical oscillators, however, can be used to easily demonstrate some of the additional levels of complexity that arise when coupling in space is considered. The Briggs-Rauscher (BR) reaction (8) is an oscillator similar to the saline oscillator in that the system continually oscillates, in this case, between two colors (see Fig. 2). Just as in the case of the SA node and the saline oscillator, the period in this system also can be changed by modifying some of the chemical constituents (11, 33), in this case, the amount of hydrogen peroxide used in the recipe. As long as the liquid remains homogenous (i.e., continually being swirled), the change in color occurs everywhere at the same time. However, if the liquid is not perturbed in this manner, small heterogeneities in the mixture will result in some regions oscillating out of phase, so that some locations will change color before others. This is the principle behind propagating waves in space, where only part of the domain is excited, and the excited state can advance by exciting the unexcited (quiescent) part of the domain.

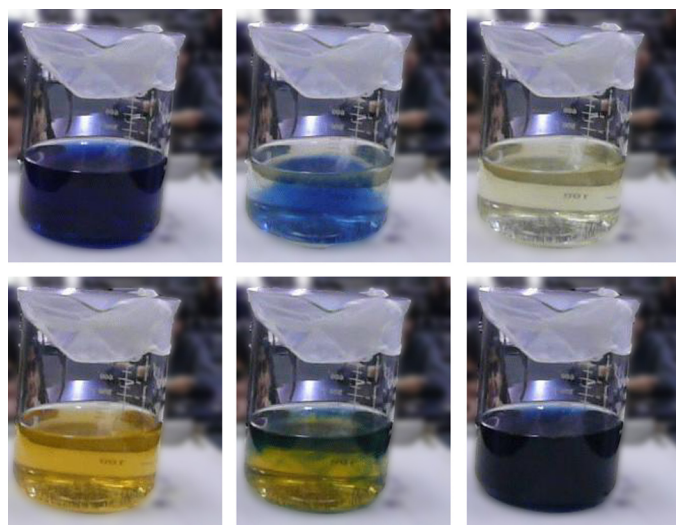


Fig. 2. The Briggs-Rauscher is a visually striking chemical reaction that oscillates from dark blue to bright yellow with a brief moment in between when it is colorless. The oscillation can last a couple of minutes, and the change in color is produced by the continuous oscillation between the concentrations of iodine and iodide ions by oxygen and carbon dioxide. The oscillation continues until the malonic acid is depleted. (For a movie of the reaction, see http://TheVirtualHeart.org/CMACS/Briggs-Rauscher_reaction1s.mov.)

To illustrate the propagation of waves, F. H. Fenton and F. B. von Stein created a set of experiments for the students using the Belousov-Zhabotinsky (BZ) reaction (6, 52, 78), which is a system that oscillates between red and blue at a much slower time scale than that of the BR reaction. In effect, the initiation and visualization of wave propagation (22, 63), as shown in Fig. 3A, is much easier. Each student was given a petri dish and instructed to pour in enough BZ solution using a pipette to form a thin (quasi-two-dimensional) layer ~1 mm deep (Fig. 3B). Students then observed the initiation of target waves due to small localized changes in concentrations. The sensitivity to initial conditions (butterfly effect) (36, 67) of chaotic systems was then explained to the students as they observed that each petri dish produced a completely different pattern even though all petri dishes were started in a similar way. They understood that even very small differences in initial conditions could make the dynamics of two instantiations diverge and produce completely different results (patterns in this case) over time. Students were encouraged to mix the petri dish many times to restart the experiment and observe the different patterns created every time.

To mimic the propagation of electrical waves in the heart, which are normally produced by the SA node or ectopic foci, students used silver wire to touch the BZ liquid (exciting the system), thereby generating propagating waves. The silver metal in the oxidizing solution forms Ag^+ , which reacts with Br^- to form the precipitate AgBr . This causes an excitation since Br^- is an inhibitor of autocatalysis (68). This variation in concentration starts to propagate by diffusion, giving rise to a propagating wave, where the size of the area of the propagating wave is a function of the diameter of the silver wire and the time that it is in contact with the solution (38).

Students then proceeded to generate spiral waves using a method similar to the initiation protocols for cardiac tissue known as the S1-S2 protocol (32) or pinwheel protocol pro-

posed by Winfree (77) modified for the BZ media (30, 38). This method generates a first wave by an initial stimulus S1 (in this case, by touching the silver wire to one section of the petri dish), after which a second stimulus S2 (again using the silver wire) is induced somewhere behind the first wave. If this second wave is at just the right distance from the back of S1, it will find a vulnerable window (65, 70) where part of the system is excitable and part is refractory, so that the second wave can propagate in only one direction, initially. As the medium immediately behind the first wave recovers, the new wave front curls, thereby initiating two counterrotating spiral waves (see Fig. 3, C and D). The generation of these spiral waves is equivalent to an induction of tachycardia in cardiac tissue, because the spiral waves have a rotation period faster than that of the natural oscillation, thereby allowing them to take control of the system at this faster period. In cardiac tissue, the natural oscillation corresponds to the activations produced by the SA node (sinus rhythm) and the faster period of tachycardia is produced by a single spiral wave or one or two pairs of counterrotating spiral waves.

The Student Research Experience

The final step of the workshop was to engage the students in a hands-on research experience. In this section, we explain the approach to engaging the students, including the elements that made this part of the workshop successful, as well as the accessibility of Java applets to develop intuition about the processes being studied and ongoing discussions about the significance of the work.

Research objective. The objective of the student-involved research was to study the dynamics of a cardiac cell model in different parameter regimes. Students were instructed that spiral waves in cardiac tissue can exhibit different dynamics depending on the physiology and structure of the tissue (2, 13, 18, 20, 25, 46, 47, 76), similar to the earlier laboratory exercise with the BZ reaction (5, 7, 49, 54). Spiral waves can rotate around small obstacles or have a very small radius of rotation

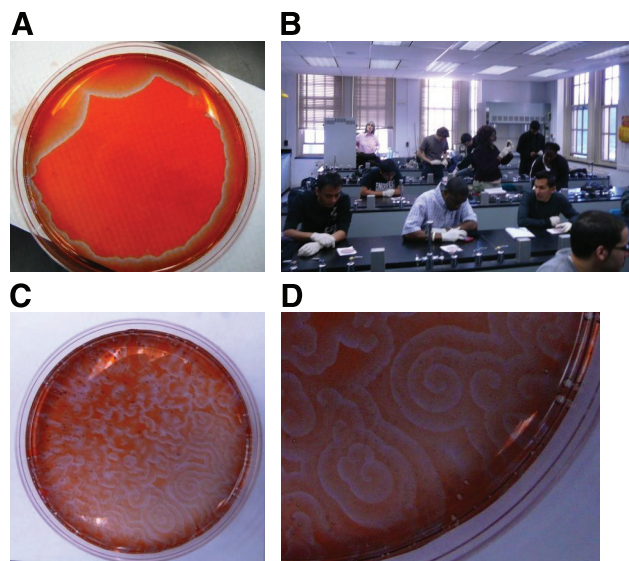


Fig. 3. Belousov-Zhabotinsky experiments. A: wave formation and propagation. B: students working on initiating spiral waves. C: example of spiral waves generated by the students. D: closeup of the spirals in C.

(see Fig. 4A), or they can have a long linear core trajectory (see Fig. 4B). In many cases, initiated spiral waves are unstable and quickly break up into multiple spiral waves (see Fig. 4C). In between these limits, there is a wide range of other possible trajectories. For chemical reactions such as the BZ reaction and for different mathematical models of cardiac cells, there have been a number of studies aimed at characterizing spiral wave dynamics as a function of model parameters (2, 13, 18, 20, 25, 46, 47, 76) to establish which physiological changes are more or less proarrhythmic.

Student preparation. During the first 2 wk, the lectures on excitable systems by F. H. Fenton and the laboratory experiments with saline oscillators and the BZ reaction gave the students a basic understanding of how the dynamics of complex systems can be modeled mathematically. As part of the early exercises, students were given time to work with a series of interactive Java applets (24) to explore the dynamics of various cardiac cell models in a single cell and in tissue (<http://TheVirtualHeart.org/applets.html>) and were encouraged to use the interactive Java applet for the simulation model in a single cell. Tutorials on numerical integration by K. Zhao and CUDA programming by J. Rogers prepared the students for the implementation of the cardiac cell model in CUDA by E. Bartocci, E. M. Cherry, and F. M. Fenton.

Group composition. In the third week, students were divided into five groups. Each group contained three students and included at least one student with courses in biology and one student with courses in computer science. Almost all of the students had taken calculus, so that was not a criterion. The objective for each group was to characterize the dynamics of spiral waves as a function of different physiological parameters to understand the effects of physiology on the simulated arrhythmias.

Methods. Students used the CUDA program downloadable from <http://TheVirtualHeart.org/CMACS/4vmCUDA.tar> to simulate the dynamics for many different values of the parameters. The *in silico* model used for this study is a reduced ionic model that reproduced the AP of a human epicardial cell (9), including experimentally measured AP shape, upstroke amplitude, and rate of rise, threshold of excitation, adaptation to changes in cycle length (i.e., the restitution curve), and conduction velocity. Briefly, in this model, the cardiac AP is modeled by three main currents: a fast inward current responsible for depolarization (positive change in voltage) that can be associated with

the fast Na^+ current, a slow outward current that is responsible for repolarization (negative change of voltage) and is analogous to a time-dependent K^+ current, and a slow inward current analogous to a total Ca^{2+} current that balances the slow outward current after depolarization to produce the plateau of the AP. Some of the model parameters can be interpreted as analogs of components of other cardiac electrophysiology models of comparable complexity, such as the Beeler-Reuter (4) and Luo-Rudy I models (50). An interactive Java applet for this model in a single cell that allows variation of any of the 28 parameters of the model to analyze how the AP characteristics change as a function of the different parameter values, including those studied by the students and presented below, can be run from <http://TheVirtualHeart.org/java/4vn/fourv0d.html>.

In each group, students initiated a spiral wave in the computational model in a manner similar to that used in the BZ reaction experiments (i.e., by an S1-S2 premature stimulation) and changed two specific sets of parameters to characterize the resulting dynamics and construct a parameter space plot. The parameters and their values are shown in Table 1. A total of 212 simulations were performed, but because the code ran essentially in real time (3), students were able to observe in a fast, easy, and interactive way how the dynamics of the reentrant waves varied as a result of parameter changes. The CUDA code is implemented with a tip-finding algorithm (27) that plots the trajectory of the spiral wave tips along with the spiral wave as the spiral evolves in time. The numerical integration used conforms with the benchmark of cardiac cell modeling (56); however, as the parameters are changed and the upstroke becomes steeper or the AP becomes very short, larger temporal or spatial resolution is required, respectively. The temporal resolution was sufficient for all the cases studied. In most cases, we used a standard five-point discretization of the Laplacian. However, for values of the time constant of the slow outward current during repolarization ($\tau_{\text{so1}} < 10$), this combination of discretization scheme and spatial resolution is insufficient and can lead to pinning to the lattice and the appearance of corners within the spiral waves (16). To ensure consistency in our comparisons, we opted to keep the same domain size fixed for all simulations and for $\tau_{\text{so1}} < 10$ instead used a nine-point Laplacian discretization whose leading error term is rotationally symmetric and thus minimizes lattice artifacts (16, 56).

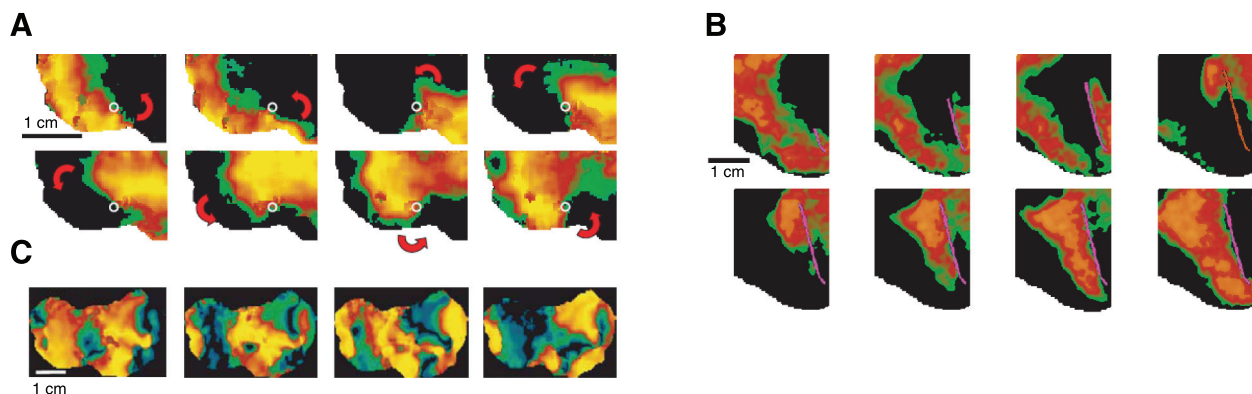


Fig. 4. Spiral waves in cardiac tissue obtained using optical mapping. A: circular core. B: linear core. C: multiple spiral wave breakup. [Used with permission from Ref. 15.]

Table 1. Summary of student group projects

Parameter	Description	Parameter Values	Number of Runs	Figure
Group 1				
τ_w^+	Time constant to inactivate the slow inward Ca^{2+} current	50, 100, 150, 200, 250, 300, 350, 400	64	5
τ_w^-	Time constant to recover from inactivation of the Ca^{2+} current	10, 25, 50, 75, 100, 150, 300, 400		
Group 2				
τ_{si}	Time constant of the slow inward Ca^{2+} current	0.05, 0.075, 0.1, 0.15, 0.2, 0.25	42	6
τ_{fi}	Time constant of the fast inward K^+ current	1.5, 1.75, 2, 2.25, 2.5, 2.75, 3.0		
Group 3				
τ_{so1}	Time constant of the slow outward current during repolarizaiton	5, 10, 15, 20, 40	35	7
τ_{si}	Time constant of the slow inward Ca^{2+} current	1.5, 1.75, 2, 2.25, 2.5, 2.75, 3.0		
Group 4				
τ_{so1}	Time constant of the slow outward current during repolarizaiton	5, 10, 15, 20, 50	35	8
τ_{fi}	Time constant of the fast inward K^+ current	0.05, 0.075, 0.1, 0.15, 0.2, 0.25, 0.3		
Group 5				
τ_{w1}^-	Time constant from inactivation of the slow inward Ca^{2+} current (first part)	5, 10, 30, 50, 150, 250	36	9
τ_{si2}	Time constant of slow inward Ca^{2+} current (second part)	15, 50, 75, 85, 100, 150		

Student Results

Students spent 5 days preparing and running simulations. Intermittently, F. H. Fenton engaged in discussion with the students to explain the results as they developed. Explanations are included below with each group's results. We believe that this kind of interaction with the students emulates the progress of a professional research project and is important to maintaining their enthusiasm for the work.

Group 1 varied the following parameters: the time constant to inactivate the slow inward Ca^{2+} current (τ_w^+) and the time constant to recover from the inactivation of the slow inward Ca^{2+} current (τ_w^-). Both parameters affect the dynamics of the inactivation gate variable w , which for the model used (9) is analogous to the voltage-dependent inactivation gate variable (f) in the Beeler-Reuter and Luo-Rudy I models. However, for this model, the time constant for inactivation is a step function, with τ_w^+ active during depolarized potentials and τ_w^- active during polarized potentials, which can be interpreted as the time constant to inactivate τ_w^+ and the time constant to recover from inactivation τ_w^- of the Ca^{2+} current. As the students could see from the Java applet, an increase (decrease) in τ_w^+ effectively increases (decreases) the AP duration (APD). The parameter τ_w^- , on the other hand, dictates the time it takes after an activation for a subsequent APD to reach the maximum APD, thereby affecting restitution properties (10, 14, 19, 53, 57, 61), where the larger the time constant, the longer it will take to reach the maximum APD.

Figure 5A shows the spiral wave profiles for the 64 different sets of parameters, and Fig. 5B shows their corresponding tip trajectories. Note that for better visualization of the trajectories, they all have been normalized to fit the same area; the real size can be estimated by observing the spiral waves. For small values of τ_w^+ , the fast inactivation of the slow inward Ca^{2+} current results, as expected, in short wavelengths, because the Ca^{2+} current is decreased, thereby reducing the plateau and leading to short APDs. As τ_w^+ increases, inactivation takes longer, resulting in increased Ca^{2+} current that produces longer APDs and longer wavelengths that yield thicker spiral

waves. A clear transition can be seen between $\tau_w^+ = 150$ ms and $\tau_w^+ = 200$ ms; in this transition, the spiral wave trajectory changes from hypocycloidal to linear.

As τ_w^- increases, the time to recover from inactivation increases, which results in a large dispersion when the APD becomes longer (i.e., large values of τ_w^+). Increased dispersion is known to be proarrhythmic (1, 17, 60, 74), in this case, by alternans (a rhythm previously shown to the students in the saline oscillator). In fact, as shown in Fig. 5A, breakup of spiral waves is obtained for larger values of both time constants. Figure 5B shows this region, where no lasting single spiral wave trajectory can be drawn, in dark gray, whereas the lighter gray colors indicate the transition region between stable and unstable spiral waves where quasi-breakup (transient breakup of a single spiral wave) occurs.

Group 2 varied the following parameters: the time constant for the slow inward Ca^{2+} current (τ_{si}) and the time constant for the Na^+ or fast inward current (τ_{fi}). The parameter τ_{si} is the inverse of the maximum Ca^{2+} conductance or slow inward current, analogous to $1/g_{si}$ in the Beeler-Reuter and Luo-Rudy I models, and τ_{fi} is the inverse of the maximum conductance for the Na^+ or fast inward current, analogous to $1/g_{Na}$ in the Beeler-Reuter and Luo-Rudy I models. The larger (smaller) τ_{si} , the smaller (larger) the Ca^{2+} current and the shorter (longer) the APD; the larger (smaller) τ_{fi} , the lower (higher) the excitability of the tissue, as the amount of Na^+ current entering the cell during depolarization decreases (increases). Again, students could verify these behaviors using the Java applet.

Figure 6 shows that the larger the APD or wavelength ($\tau_{si} < 2.0$), the longer the spiral wave tip needs to travel before finding excitable tissue to make a turn, resulting in long linear cores (21, 47). As excitability decreases, the tip takes longer to make a turn, resulting in more precession in the linear core trajectory. As τ_{si} increases, the wavelength decreases, and the spiral wave meanders more for low excitability, but as the excitability increases (smaller τ_{fi}), the faster turns make the tip of the spiral collide with its own wave back, producing quasi-breakup. Figure 6B shows the different levels of quasi-breakup

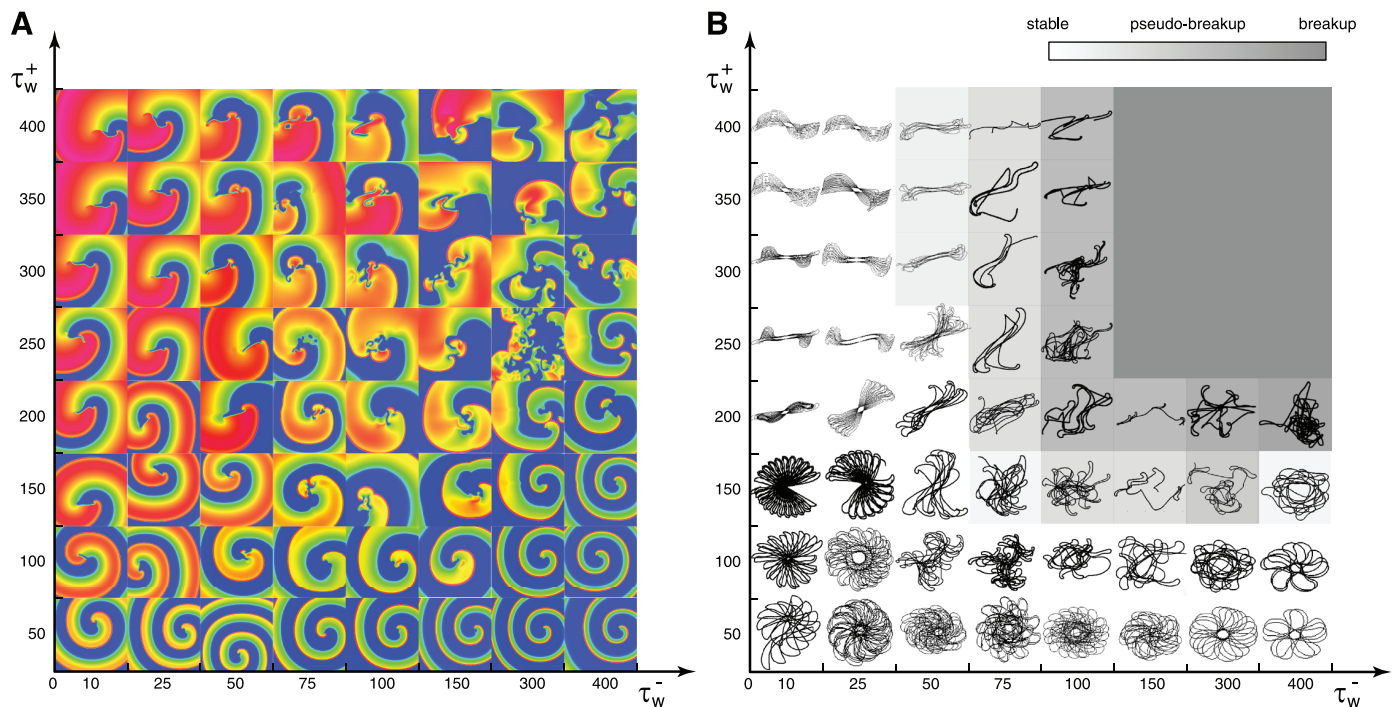


Fig. 5. Spiral wave snapshots (A) and tip trajectories (B) for combinations of the following parameters: the time constant to inactivate the slow inward Ca^{2+} current (τ_w^+) and the time constant to recover from the inactivation of the slow inward Ca^{2+} current (τ_w^-). Instability is promoted by increasing the values of both parameters, leading to quasi-breakup or full breakup (gray shaded regions in the tip trajectory plot). See text for details.

and meander as a function of the wavelength (determined here by τ_{si}) and excitability (determined here by τ_{fi}) of the simulated tissue.

Group 3 varied the following parameters: τ_{so1} and τ_{si} (described above). The parameter τ_{so1} is the inverse of the maximum conductance of the slow outward current during depolarization, so it is similar to $1/\bar{g}_{X1}$ in the Beeler-Reuter model and $1/G_KX_1$ in the Luo-Rudy I model; τ_{si} is as described above.

The larger (smaller) τ_{so1} , the smaller (larger) the effective K^+ current and the larger (smaller) the APD. For small values of τ_{si} , the repolarization current is so strong that the APD becomes as small as that of a mouse or rat, resulting in very thin spiral waves similar to those obtained with the BZ reaction, as shown in Fig. 7. As the repolarization current decreases (larger τ_{so1}), the AP plateau duration increases, and the wavelength of the spiral increases. As the Ca^{2+} current increases

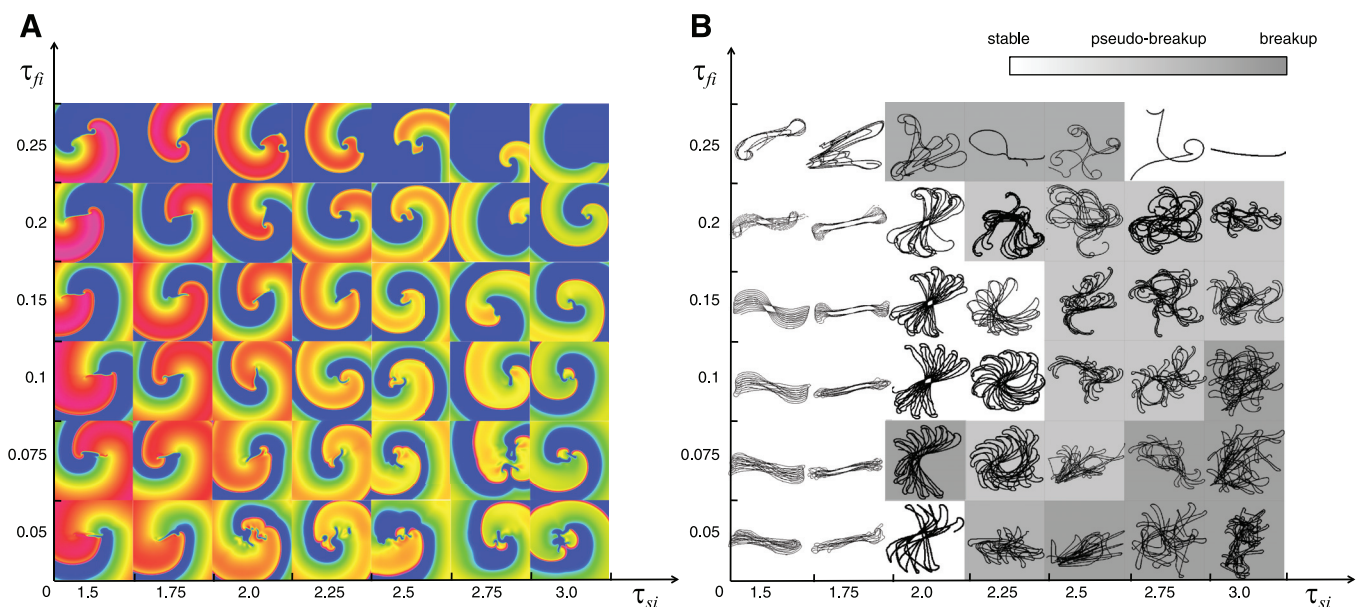


Fig. 6. Spiral wave snapshots (A) and tip trajectories (B) for combinations of the following parameters: the time constant for the slow inward Ca^{2+} current (τ_{si}) and the time constant for the Na^+ or fast inward current (τ_{fi}). Instability is promoted by increasing the value of τ_{si} , leading, in most cases, to quasi-breakup or full breakup (gray shaded regions in the tip trajectory plot). See text for details.

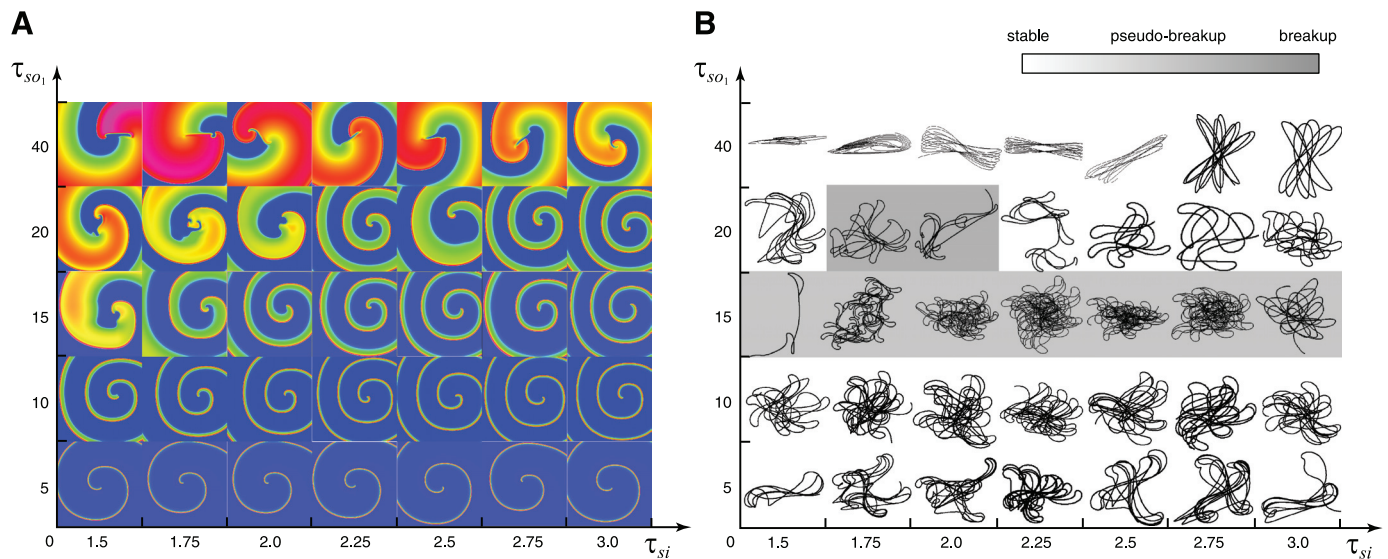


Fig. 7. Spiral wave snapshots (A) and tip trajectories (B) for combinations of the following parameters: the time constant of the slow outward current during depolarization (τ_{so1}) and τ_{si} . Instability is strongest for intermediate values of τ_{so1} (gray shaded regions in the tip trajectory plot). See text for details.

(smaller values of τ_{si}), the tip meanders less, and the linear cores precess less. Figure 7 shows that for these sets of parameter values, intermediate values of the repolarization current (τ_{so1}) produce a quasi-breakup regime. This occurs in the intermediate regime in between meander and linear cores, where complex hypermeandering trajectories are present. When hypermeander is complex, in some cases, the spiral wave tip can turn so fast that a Doppler shift effectively increases the frequency of the spiral wave locally to the point that it reaches the refractory period, producing temporary (nonsustained) breakup that subsequently heals.

Group 4 varied the following parameters: τ_{so1} and τ_{fi} (both described previously). Figure 8 shows that as the excitability decreases and the repolarization current is quite strong, no spiral waves can be induced because there is not enough electrotonic current to produce an excitation to sustain wave

propagation. As in the previous case, a large repolarization current results in spiral waves with small wavelengths; however, as the excitability increases (smaller τ_{fi}), the tip of the spiral wave can turn faster, as it needs to excite fewer neighbors to propagate (28, 47), thereby resulting in a faster rotation frequency and tighter spiral waves compared with those shown in Fig. 7. However, as with the results from group 3, there are values of depolarization and hypermeandering trajectories for which transient breakup by the Doppler effect occurs.

Group 5 varied the following parameters: the time constant from inactivation of the slow inward Ca^{2+} current (first part) (τ_{w1}^-) and the time constant of slow inward Ca^{2+} current (second part) (τ_{si2}). As in group 1, τ_{w1}^- represents the recovery from inactivation for the slow inward current, and, as in groups 2 and 3, τ_{si} is the time constant for the slow inward Ca^{2+} current. In the minimal model (9), these parameters can be

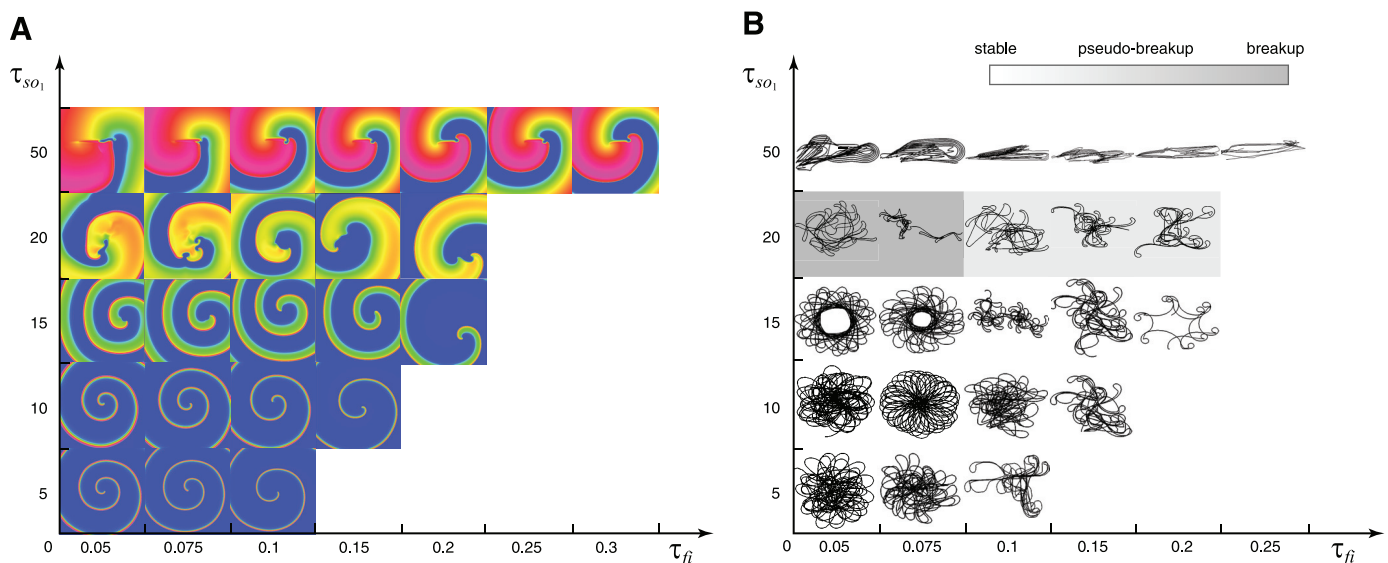


Fig. 8. Spiral wave snapshots (A) and tip trajectories (B) for combinations of the following parameters: τ_{so1} and τ_{fi} . Instability is strongest for intermediate values of τ_{so1} (gray shaded regions in the tip trajectory plot). See text for details.

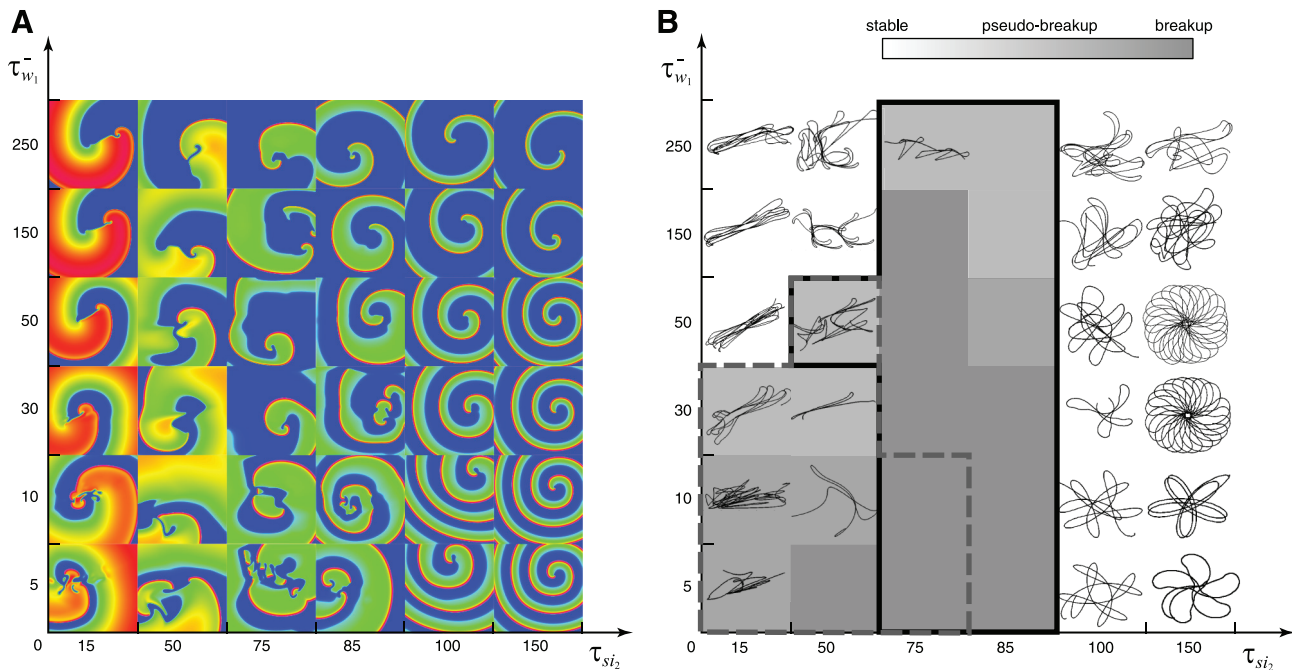


Fig. 9. Spiral wave snapshots (A) and tip trajectories (B) for combinations of the following parameters: the time constant from inactivation of the slow inward Ca^{2+} current (first part) (τ_{w1}) and the time constant of slow inward Ca^{2+} current (second part) (τ_{si2}). Instability is promoted primarily by decreasing the value of τ_{si} and, when τ_{si} is small, by decreasing τ_{w1} as well. See text for details.

piece-wise functions between two values (e.g., τ_{w1}^- and τ_{w2}^- , and τ_{si1} and τ_{si2} , respectively); in this case, only τ_{w1}^- and τ_{si2} were varied while keeping the other two values constant at the values given in Ref. 9. τ_{w1}^- can produce large regions of dispersion when the APD is long, as in the case of small values of τ_{si2} . Because no parameter related to the upstroke (fast inward current) of the model has been changed, the excitability remains relatively high, resulting in relatively thick spiral waves (see Fig. 9A) that exhibit fast turns. As shown in Fig. 9B, there is a large region of spiral wave breakup and quasi-breakup (shown in grayscale). In this case, the mechanisms described in Fig. 5 and those in Figs. 6–8 are mixed. Most of the large dispersion and alternans breakup occurs within the region delimited by the solid line, whereas the Doppler effect breakup occurs within the region delimited by the dashed line.

Conclusions

The goals of the workshop were to cultivate student interest in the area of computational modeling and analysis of complex systems and to provide students early in their academic careers with a hands-on research experience. This article is itself evidence of their engagement in a hands-on research experience, and student feedback, as shown in Table 2, indicated that student interest was captured effectively.

Using simpler systems such as the saline oscillator and chemical oscillators, the students not only gained first-hand experience in experimental studies but also worked individually and as teams to answer questions that helped them understand the dynamics of complex systems and to participate actively in a research study that resulted in this report on which they appeared as co-authors. The experimental component therefore prepared them to develop a stronger understanding of how cardiac cells work, to learn how mathematical models are

used for the study of arrhythmias, and to study how changes in physiological properties can alter the electrical dynamics of cardiac cells in tissue. From the five independent group studies, the students have shown that spiral wave dynamics and trajectories depend on model parameters. As has been observed in other models (2, 13, 18, 20, 25, 46, 47, 76), as the wavelength increases, two main effects can be seen: the likelihood of linear core trajectories increases and breakup becomes increasingly probable. Conversely, as the wavelength decreases, the likelihood of breakup decreases; however, the range of possible tip trajectories becomes much broader. In one particularly novel finding, students were able to demonstrate for the first time that, although previous studies of spiral waves with linear cores have always shown a precession that makes the angle of rotation vary, it is possible to obtain spiral waves with linear cores that do not precess for particular regions in parameter space with certain pairs of values for τ_w^+ and τ_w^- (see Fig. 5B) or τ_{fi} and τ_{si} (see Fig. 6B).

Table 2. Student feedback to various questions

	Median	Average
The workshop helped me to understand how to analyze and use models of biological processes.	4.5	4.3
The workshop helped me to understand how to construct and use models of biological processes.	4	3.8
The workshop helped me to understand how to formulate and examine scientific hypotheses.	4	4
I feel more confident of my ability to do scientific or technical work as a result of attending the workshop.	5	4.6
The workshop made me more interested in continuing my schooling in a scientific, technical, engineering, or mathematical field.	5	4.9

$n = 15$ student respondents. Questions were scored on a scale from 1 (disagree completely) to 5 (agree completely).

We anticipate that some of the students from the workshop will continue working with the researchers leading the workshop over the summer, a result that also occurred in the last workshop, where two students obtained summer research jobs at Carnegie Mellon University.

APPENDIX

The BR Reaction

A 50-ml volume containing sodium iodate (328.5 mM) and sulfuric acid (130 mM) in water was mixed with an equal volume of a solution containing malonic acid (200 mM), manganese (II) sulfate monohydrate (26 mM), and soluble starch (2.5 mM) in water. A 100-ml volume of 3% hydrogen peroxide was then added to create the reaction.

The BZ Reaction

A 50-ml volume containing sodium bromate (250 mM) in water was mixed with an equal volume of malonic acid (310 mM) and sodium bromide (58 mM) in water. After the solution cleared, a 50-ml volume of cerium (IV) ammonium nitrate (20 mM), sulfuric acid (2.7 M), and 3 ml ferroin (25 mM) was added to create the reaction.

The BZ Reaction for Waves

Seven milliliters of sodium bromate (0.5 M) in sulfuric acid (0.6 M) were mixed with 3.5 ml of malonic acid (0.5 M) in water and 1 ml of sodium bromide (0.97 M) in water. After the solution cleared, 1 ml of ferroin (25 mM) was added, and the solution was again mixed. A volume of this solution was transferred to a clean petri dish to form a thin layer 1–2 mm deep to observe spontaneous spiral wave formation.

Neutralization of the Chemical Reactions

BR reaction. Neutralize the reduced iodide with the addition of 5 g sodium thiosulfate per liter of demonstration solution. Flush down the drain with water.

BZ reaction. Neutralize with sodium bicarbonate. Flush down the drain with plenty of water.

CUDA Program

A copy of the CUDA code used by the students in this article can be downloaded from <http://TheVirtualHeart.org/CMACS/4vmCUDA>. To run it, one needs a computer with an NVIDIA card. You can download the CUDA drivers and toolkit from the NVIDIA web site: <http://developer.nvidia.com/cuda-toolkit-40>.

Install the CUDA drivers.

Install the CUDA toolkit.

Make ./cuda2DMinModel.

Below is a shortcut of the keyboard functions that can be used during simulations:

- “P” key: pause the simulation
- “R” key: restart the simulation
- “Q” key: quit the simulation
- “U” key: show the u field
- “V” key: show the v field
- “W” key: show the w field
- “S” key: show the s field
- “T” key: show or hide the tip trajectory

ACKNOWLEDGMENTS

The authors thank Humberto Arce and Hortencia Gonzales for the helpful advice in preparing the saline oscillator setups and Kenneth Showalter for comments on the chemical section. The authors also thank the Information Technology Department of Lehman College for access to the computer

laboratory where most of the workshop took place and Brian Murphy for the use of CUDA-ready computers.

GRANTS

This work was supported by National Science Foundation Grants CCF-0926190 and CCF-1018459 and in part by Air Force Office of Scientific Research Grant FA0550-09-1-0481.

DISCLOSURES

No conflicts of interest, financial or otherwise, are declared by the author(s).

REFERENCES

1. **Banville I, Gray RA.** Effect of action potential duration and conduction velocity restitution and their spatial dispersion on alternans and the stability of arrhythmias. *J Cardiovasc Electrophysiol* 13: 1141–1149, 2002.
2. **Barkley D.** Euclidean symmetry and the dynamics of rotating spiral waves. *Phys Rev Lett* 72: 164, 1994.
3. **Bartocci E, Cherry EM, Glimm J, Grosu R, Smolka SA, Fenton FH.** Toward real-time simulation of cardiac dynamics. In: *Proceedings of the 9th International Conference on Computational Methods in Systems Biology*. New York: ACM, 2011, p. 103–110.
4. **Beeler GW, Reuter H.** Reconstruction of the action potential of ventricular myocardial fibres. *J Physiol* 268: 177–210, 1977.
5. **Belmonte AL, Ouyang Q, Flesselles JM.** Experimental survey of spiral dynamics in the Belousov-Zhabotinsky reaction. *J Physique II* 7: 44, 1997.
6. **Belousov BP.** A periodic reaction and its mechanism. *Compil Abstr Radiat Med*: 147, 1959.
7. **Braune M, Engel H.** Compound rotation of spiral waves in a light-sensitive Belousov-Zhabotinsky medium. *Chem Phys Lett* 204: 257–264, 1993.
8. **Briggs TS, Rauscher WC.** An oscillating iodine clock. *J Chem Educ* 50: 496, 1973.
9. **Bueno-Orovio A, Cherry EM, Fenton FH.** Minimal model for human ventricular action potentials in tissue. *J Theor Biol* 253: 544–60, 2008.
10. **Cao JM, Qu Z, Kim YH, Wu TJ, Garfinkel A, Weiss JN, Karagueuzian HS, Chen PS.** Spatiotemporal heterogeneity in the induction of ventricular fibrillation by rapid pacing: importance of cardiac restitution properties. *Circ Res* 84: 1318–1331, 1999.
11. **Cervellati R, Höner K, Furrow SD, Neddens C, Costa S.** The Briggs-Rauscher reaction as a test to measure the activity of antioxidants. *Helv Chim Acta* 84: 3533–3547, 2001.
12. **Cervellati R, Soldà R.** An alternating voltage battery with two salt-water oscillators. *Am J Phys* 69: 543, 2001.
13. **Cherry EM, Evans SJ.** Properties of two human atrial cell models in tissue: restitution, memory, propagation, and reentry. *J Theor Biol* 254: 674–90, 2008.
14. **Cherry EM, Fenton FH.** Suppression of alternans and conduction blocks despite steep APD restitution: electrotonic, memory, and conduction velocity restitution effects. *Am J Physiol Heart Circ Physiol* 286: H2332–H2341, 2004.
15. **Cherry EM, Fenton FH.** Visualization of spiral and scroll waves in simulated and experimental cardiac tissue. *N J Phys* 10: 125016, 2008.
16. **Clayton RH, Bernus O, Cherry EM, Dierckx H, Fenton FH, Mirabella L, Panfilov AV, Sachse FB, Seemann G, Zhang H.** Models of cardiac tissue electrophysiology: progress, challenges and open questions. *Prog Biophys Mol Biol* 104: 22–48, 2011.
17. **Clayton R, Taggart P.** Regional differences in APD restitution can initiate wavebreak and re-entry in cardiac tissue: a computational study. *Biomed Eng Online* 4: 54, 2005.
18. **Comtois P, Kneller J, Nattel S.** Of circles and spirals: bridging the gap between the leading circle and spiral wave concepts of cardiac reentry. *Europace* 7: S10–S20, 2005.
19. **Courtemanche M.** Complex spiral wave dynamics in a spatially distributed ionic model of cardiac electrical activity. *Chaos* 6: 579–600, 1996.
20. **Davidenko JM, Pertsov AV, Salomonsz R, Baxter W, Jalife J.** Stationary and drifting spiral waves of excitation in isolated cardiac muscle. *Nature* 355: 349–51, 1992.
21. **Efimov IR, Krinsky VI, Jalife J.** Dynamics of rotating vortices in the Beeler-Reuter model of cardiac tissue. *Chaos Solitons Fractals* 5: 513–526, 1995.
22. **Epstein IR, Pojman JA.** *An Introduction to Nonlinear Chemical Dynamics: Oscillations, Waves, Patterns, and Chaos*. New York: Oxford Univ. Press, 1998.

23. Fenton FH, Cherry EM, Glass L. Cardiac arrhythmia. *Scholarpedia* 3: 1665, 2008.
24. Fenton FH, Cherry EM, Hastings HM, Evans SJ. Real-time computer simulations of excitable media: JAVA as a scientific language and as a wrapper for C and FORTRAN programs. *Biosystems* 64: 73–96, 2002.
25. Fenton FH, Cherry EM, Hastings HM, Evans SJ. Multiple mechanisms of spiral wave breakup in a model of cardiac electrical activity. *Chaos* 12: 852–892, 2002.
26. Fenton FH, Cherry EM. Models of cardiac cell. *Scholarpedia* 3: 1868, 2008.
27. Fenton FH, Karma A. Vortex dynamics in three-dimensional continuous myocardium with fiber rotation: filament instability and fibrillation. *Chaos* 8: 20–47, 1998.
28. Fenton FH, Evans SJ, Hastings HM. Memory in an excitable medium: a mechanism for spiral wave breakup in the low-excitability limit. *Phys Rev Lett* 83: 3964, 1999.
30. Fernandez-Garcia G, Gomez-Gesteira M, Munuzuri AP, Perez-Munuzuri V, Perez-Villar V. A method for spiral wave generation in the Belousov-Zhabotinsky reaction. *Eur J Phys* 15: 221–227, 1994.
31. Fink M, Niederer SA, Cherry EM, Fenton FH, Koivumäki JT, Seemann G, Thul R, Zhang H, Sachse FB, Beard D, Crampin EJ, Smith NP. Cardiac cell modelling: observations from the heart of the cardiac physiome project. *Prog Biophys Mol Biol* 104: 2–21, 2011.
32. Frazier DW, Wolf PD, Wharton JM, Tang AS, Smith WM, Ideker RE. Stimulus-induced critical point. Mechanism for electrical initiation of reentry in normal canine myocardium. *J Clin Invest* 83: 1039–1052, 1989.
33. Furrow SD, Cervellati R, Amadori G. New substrates for the oscillating Briggs-Rauscher reaction. *J Phys Chem A* 106: 5841–5850, 2002.
34. Glass L, Guevara MR, Shrier A. Universal bifurcations and the classification of cardiac arrhythmias. *Ann NY Acad Sci* 504: 168–178, 1987.
35. Glass L, Mackey MC. *From Clocks to Chaos*. Princeton, NJ: Princeton Univ. Press, 1988.
36. Gleick J. *Chaos: Making a New Science*. New York: Penguin, 1988.
37. Glimm J, Grove JW, Li XL, Oh W, Sharp DH. A critical analysis of Rayleigh-Taylor growth rates. *J Comp Phys* 169: 652–677, 2001.
38. Gómez-Gesteira M, Fernández-García G, Muñozuri AP, Pérez-Munuzuri V, Krinsky VI, Starmer CF, Pérez-Villar V. Vulnerability in excitable Belousov-Zhabotinsky medium: from 1D to 2D. *Physica D* 76: 359–368, 1994.
39. González H, Arce H, Guevara MR. Phase resetting, phase locking, and bistability in the periodically driven saline oscillator: Experiment and model. *Phys Rev E* 78: 036217, 2008.
40. Guevara MR, Ward G, Shrier A, Glass L. Electrical alternans and period-doubling bifurcations. *Comp Cardiol* 1984: 167–170, 1984.
41. Hodgkin AL, Huxley AF. A quantitative description of membrane currents and its application to conduction and excitation in nerve. *J Physiol* 117: 500–544, 1952.
42. Huffaker CB. Experimental studies on predation: dispersion factors and predator-prey oscillations. *Hilgardia* 27: 343–383, 1958.
43. Izhikevich E, FitzHugh R. FitzHugh-Nagumo model. *Scholarpedia* 1: 1349, 2006.
44. Kano T. *Experimental and Theoretical Study on Density Oscillator*. (PhD Thesis). Osaka: Osaka Univ., 2008.
45. Karafyllidis I, Thanailakis A. A model for predicting forest fire spreading using cellular automata. *Ecol Model* 16: 87–97, 1997.
46. Kim DT, Kwan Y, Lee JJ, Ikeda T, Uchida T, Kamjoo K, Kim YH, Ong JJC, Athill CA, Wu TJ, Czer L, Karagueuzian HS, Chen PS. Patterns of spiral tip motion in cardiac tissues. *Chaos* 8: 137–148, 1998.
47. Krinsky VI, Efimov IR, Jalife J. Vortices with linear cores in excitable media. *Proc R Soc Lond A* 437: 645–655, 1992.
48. Lorenz EN. Deterministic nonperiodic flow. *J Atmos Sci* 20: 130–141, 1963.
49. Luengviriyi C, Storb U, Hauser MJB, Müller SC. An elegant method to study an isolated spiral wave in a thin layer of a batch Belousov-Zhabotinsky reaction under oxygen-free conditions. *Phys Chem Chem Phys* 8: 1425, 2006.
50. Luo CH, Rudy Y. A model of the ventricular cardiac action potential. Depolarization, repolarization, and their interaction. *Circ Res* 68: 1501–1526, 1991.
51. Martin S. A hydrodynamic curiosity: the salt oscillator. *Geophys Astrophys Fluid Dyn* 1: 143–160, 1970.
52. Mikhailov AS, Showalter K. Control of waves, patterns and turbulence in chemical systems. *Phys Rep* 425: 79–194, 2006.
53. Morgan JM, Cunningham D, Rowland E. Dispersion of monophasic action potential duration: demonstrable in humans after premature ventricular extrastimulation but not in steady state. *J Am Coll Cardiol* 19: 1244–1253, 1992.
54. Nagy-Ungvarai Z, Tyson JJ, Mueller SC, Watson LT, Hess B. Experimental study of spiral waves in the cerium-catalyzed Belousov-Zhabotinsky reaction. *J Phys Chem* 94: 8677–8682, 1990.
55. Nayagam V, Williams FA. Rotating spiral edge flames in von Karman swirling flows. *Phys Rev Lett* 84: 479, 2000.
56. Niederer SA, Kerfoot E, Benson A, Bernabeu M, Bernus O, Bradley C, Cherry EM, Clayton R, Fenton FH, Garny A, Heidenreich E, Land S, Maleckar M, Pathmanathan P, Planck G, Rodriguez JF, Roy I, Sachse FB, Seemann G, Skavhaug O, Smith N. Verification of cardiac tissue electrophysiology simulators using an N-version benchmark. *Phil Trans R Soc Lond A*. 369: 4331–4351, 2011.
57. Nolasco JB, Dahlen RW. A graphic method for the study of alternation in cardiac action potentials. *J Appl Physiol* 25: 191–196, 1968.
58. Noyes RM. A simple explanation of the salt water oscillator. *J Chem Educ* 66: 207, 1989.
59. Okamura M, Yoshikawa K. Rhythm in a saline oscillator. *Phys Rev E* 61: 2445, 2000.
60. Pastore JM, Girouard SD, Laurita KR, Akar FG, Rosenbaum DS. Mechanism linking T-wave alternans to the genesis of cardiac fibrillation. *Circulation* 99: 1385–1394, 1999.
61. Riccio ML, Koller ML, Gilmour RF. Electrical restitution and spatio-temporal organization during ventricular fibrillation. *Circ Res* 84: 955–63, 1999.
62. Sanders J, Kandrot E. *CUDA by Example: an Introduction to General-Purpose GPU Programming* (1st ed.). Boston, MA: Addison-Wesley Professional, 2010.
63. Shakhshiri BZ. *Chemical Demonstrations: a Handbook for Teachers of Chemistry* (1st ed.). Madison, WI: Univ. of Wisconsin Press, 1983, vol. 1.
64. Sharp DH. An overview of Rayleigh-Taylor instability. *Physica D* 12: 3–18, 1984.
65. Shaw RM, Rudy Y. The vulnerable window for unidirectional block in cardiac tissue: characterization and dependence on membrane excitability and intercellular coupling. *J Cardiovasc Electrophysiol* 6: 115–131, 1995.
66. Sherratt JA, Smith MJ. Periodic travelling waves in cyclic populations: field studies and reaction-diffusion models. *J R Soc Interface* 5: 483–505, 2008.
67. Shinbrot T, Ditto W, Grebogi C, Ott E, Spano M, Yorke JA. Using the sensitive dependence of chaos (the “butterfly effect”) to direct trajectories in an experimental chaotic system. *Phys Rev Lett* 68: 2863, 1992.
68. Showalter K, Noyes RM, Turner H. Detailed studies of trigger wave initiation and detection. *J Am Chem Soc* 101: 7463–7469, 1979.
69. Shrier A, Dubarsky H, Rosengarten M, Guevara M, Nattel S, Glass L. Prediction of complex atrioventricular conduction rhythms in humans with use of the atrioventricular nodal recovery curve. *Circulation* 76: 1196–1205, 1987.
70. Starmer C. Vulnerability of cardiac dynamics. *Scholarpedia* 2: 1847, 2007.
71. Strogatz SH. *Nonlinear Dynamics and Chaos: With Applications to Physics, Biology, Chemistry, and Engineering* (1st ed.). Boulder, CO: Westview, 2001.
72. Truscott JE, Brindley J. Ocean plankton populations as excitable media. *Bull Math Biol* 56: 981–998, 1994.
73. Upadhyay S, Das AK, Agarwala V, Srivastava RC. Oscillations of electrical potential differences in the salt-water oscillator. *Langmuir* 8: 2567–2571, 1992.
74. Watanabe MA, Fenton FH, Evans SJ, Hastings HM, Karma A. Mechanisms for discordant alternans. *J Cardiovasc Electrophysiol* 12: 196–206, 2001.
75. Wiener N, Rosenblueth A. The mathematical formulation of the problem of conduction of impulses in a network of connected excitable elements, specifically in cardiac muscle. *Arch Inst Cardiol Mex* 16: 205–265, 1946.
76. Winfree AT. Varieties of spiral wave behavior: an experimentalist’s approach to the theory of excitable media. *Chaos* 1: 303–334, 1991.
77. Winfree AT. *The Geometry of Biological Time* (2nd ed.). New York: Springer, 2001.
78. Zhabotinsky AM. Periodic processes of malonic acid oxidation in a liquid phase. *Biofizika* 9: 306–311, 1964.